

# Persistence of Long-Term Memory in Vitrified and Revived *Caenorhabditis elegans*

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## Abstract

Can memory be retained after cryopreservation? Our research has attempted to answer this long-standing question by using the nematode worm *Caenorhabditis elegans*, a well-known model organism for biological research that has generated revolutionary findings but has not been tested for memory retention after cryopreservation. Our study's goal was to test *C. elegans*' memory recall after vitrification and reviving. Using a method of sensory imprinting in the young *C. elegans*, we establish that learning acquired through olfactory cues shapes the animal's behavior and the learning is retained at the adult stage after vitrification. Our research method included olfactory imprinting with the chemical benzaldehyde (C<sub>6</sub>H<sub>5</sub>CHO) for phase-sense olfactory imprinting at the L1 stage, the fast-cooling SafeSpeed method for vitrification at the L2 stage, reviving, and a chemotaxis assay for testing memory retention of learning at the adult stage. Our results in testing memory retention after cryopreservation show that the mechanisms that regulate the odorant imprinting (a form of long-term memory) in *C. elegans* have not been modified by the process of vitrification or by slow freezing.

## Introduction

*Caenorhabditis elegans* IS ONE OF THE MOST IMPORTANT models used in biology and neurology<sup>1</sup> and has countless applications in the area of biological sciences.<sup>2-4</sup> The simplicity of its size (1 mm), the transparency of its neuronal network (hermaphrodites contain 302 neurons)<sup>5</sup> and its short but complex life cycle make *C. elegans* of potential value to studies of memory retention after cryopreservation. Through the use of *C. elegans* as a model, numerous experiments have been performed in the area of memory and neurodegenerative diseases,<sup>3,4</sup> many of which explain the mechanism of memory, including the differences between short- and long-term memory.<sup>6-13</sup>

Our study addresses the specific interest that long- or short-term memories of cryopreserved and revived animals have not been tested. The organism of choice to explore this question is *C. elegans* because it is the only organism in which both cryopreservation and revival have been demonstrated and there is a well-defined assay of learning.

To evaluate our study's specific interest, we combined and used three principal methodologies developed previously: (1) Olfactory imprinting for *C. elegans*,<sup>9</sup> (2) vitrification of *C. elegans* by the SafeSpeed cryopreservation

protocol,<sup>14</sup> and (3) a chemotaxis migration assay.<sup>9</sup> Attached to this project and as an extra goal, we also evaluated the persistence of memory using the traditional protocol of cryopreservation of *C. elegans* by slow freezing.<sup>2</sup>

Our research has established a protocol for examining the results of olfactory imprinting, demonstrating long-term memory retention of this simple adult animal after vitrification and reviving. This study provides new knowledge to the field of cryopreservation, and we consider such findings to be a preliminary step in ascertaining the preservation of memory of simple animals undergoing vitrification.

## Learning in *C. elegans*

In our study, we first explored learning through chemical attractants for testing memory of behavior performance. In diverse environments, *C. elegans* have been shown to habituate to learned smells and to associate attractant and aversive chemicals with the presence or absence of food.<sup>9,11,12</sup> One method for training and testing learning in the worms is to expose them to a specific chemical compound over a restricted period of time with the presence or absence of food to create a pattern of behavior. When encountering the chemical compound at later times, the worms' level of

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response (memory retention) is tested and evaluated by a chemotaxis migration assay.<sup>9,12</sup> In their findings of associative short-term memory, Kauffman et al.<sup>12</sup> showed that after *C. elegans* are exposed to the chemoattractant butanone, the worms' memory response starts to decrease 1 hr after exposure. In their findings on long-term memory, Remy and Hobert<sup>9</sup> showed that after *C. elegans* are exposed to attractant chemicals such as benzaldehyde at the early larval stage, the worms' memory response in the adult stage continues to be retained after 5 days, or 120 hr, for phase-sense learning, referred to as imprinting.

Imprinting is one of the mechanisms key to analyzing basic neural aspects of memory<sup>15</sup> and is known as a type of learning in which young organisms make strong associations. Because of the potential for strong associations, imprinting is often used to explain early adaptation or bonding to particular sensory cues, such as olfactory signals. Olfactory-based imprinting has been studied in many species, including humans, other primates, mammals, ungulates, aquatic animals, birds, and microscopic simple animals (such as *C. elegans*). In olfactory imprinting,<sup>9</sup> there is a distinct period of time (a window of opportunity or physiological state that lasts for a limited time) in which an organism can develop a long-lasting learned response. Notably, the distinct period of time distinguishes this form of memory from other types of learned olfactory adaptive procedures in *C. elegans* that only last for a limited time and in which olfactory imprinting behavior is mediated by a single pair of interneurons.<sup>9</sup> Moreover, there is evidence that olfactory imprinting of *C. elegans* is transmitted to the next generation if the imprint is accomplished successfully for more than four generations.<sup>16</sup>

### Cryopreservation of *C. elegans*

There are two traditional ways to cryopreserve biological samples: (1) Slow freezing and (2) vitrification.<sup>17,18</sup> These cryopreservation methods are differentiated by the cooling and warming rates and the concentration of cryoprotectant used to obtain successful cryopreservation, whether or not ice crystals are formed, and the attendant dehydration of the cells.

Slow freezing requires a low concentration of cryoprotectant and slow cooling and warming rates with the formation of ice crystals. Although this method has been the traditional protocol for cryopreserving *C. elegans*, it has a limited survival rate of around 25%–35% for the L1 and L2 stages.<sup>2</sup>

Vitrification is the transformation of a liquid substance into a vitreous solution, achieved by high cooling rates and concentrations of cryoprotectant, eliminating ice formation.<sup>18–20</sup> The basic method for vitrifying cells is known as Cryotop, which has been used for embryos and oocytes for over a decade with excellent results on the basis of survival rates and is considered to be the most reliable cryopreservation method for cells and embryos.<sup>21,22</sup> The Cryotop vitrification protocol directly submerges cells into liquid nitrogen. This method developed from minimum volume cooling (MVC)<sup>23</sup> using traditional sample straws<sup>24</sup> and proved to have a higher survival rate for cells as compared to conventional vitrification, due to the much higher cooling rates attainable with small volumes.

Combining the thermal performance of quartz capillaries, ultra-fast cooling-rates (slush nitrogen),<sup>19,25,26</sup> and MVC,<sup>22</sup>

the protocol for vitrification of *C. elegans* in the L1 and L2 stages was developed.<sup>26</sup> However, the warming rate in this protocol was too slow to successfully achieve vitrification of other stages of the larvae and the adult. Recent studies concerning the importance of the warming rates, more than the cooling rates,<sup>27–30</sup> have been contributing factors in developing new techniques to improve vitrification protocols.

The SafeSpeed closed device<sup>31</sup> is a new technology based on ultra-fast warming rates that was originally developed to revive vitrified oocytes and embryos and offers an alternative for vitrifying *C. elegans*, simplifying the protocol and ensuring survival rates ~100% for all *C. elegans* stages, including the adult stage.<sup>14</sup> This method also presents an excellent alternative for cryopreserving *C. elegans*, with much better survival rates (for all stages) than slow freezing and is an easier protocol than vitrification using quartz capillaries.<sup>14,25</sup>

## Materials and Methods

### *C. elegans* native strain

Worm maintenance, including breeding, was conducted as described by Stiernagle.<sup>32</sup> The Wild isolate Bristol strain N2 was obtained from the Caenorhabditis Genetics Center (CGC) at the University of Minnesota.

### Control group

The control group consisted of eight sets of 100 or more worms for each study. We used a combination of variables in each study to observe and identify the native behavior of the worms in the chemotaxis assay and the effects of cryoprotectant solutions and cryopreservation protocols on the worms' behavior. The eight sets of studies are identified as S1, S2, S3, S4, S5, S7, S8, and S9 (Table 1).

We applied the term "untrained" to sets of worms that were not imprinted with benzaldehyde and the term "trained" to sets of worms that were imprinted with benzaldehyde. Studies included assembling of trained and untrained worms and the presence or absence of a cryopreservation protocol. First, trained and untrained worms were exposed to either the cryoprotectant vitrification solution or the cryoprotectant slow freezing solution; and, second, the cryopreservation protocols were completed by vitrification or by slow freezing (Table 1).

### Experimental group

We describe the experimental group as two sets of 100 or more worms for each study. The worms were trained, cryopreserved, revived, and tested for memory recall. The cryopreservation methods included worms that were trained and cryopreserved by vitrification and worms that were trained and cryopreserved by slow freezing. This was the most important study to test the retention of memory after vitrification or slow freezing. The two sets of studies are identified as S6 and S10 (Table 1).

### Phase-sense olfactory imprinting

Using the Remy and Hobert<sup>9</sup> method for olfactory imprinting, 20 L1 stage worms were pipetted into a 6-cm petri dish with a lawn of *Escherichia coli* OP50 and exposed to

TABLE 1. DESCRIPTION OF THE STUDIES

	Olfactory imprinting		Cryoprotectant solutions		Cryopreservation	
	Trained (benzaldehyde)	Untrained (distilled water)	Vitrification solution	Slow freezing solution	Vitrification	Slow freezing
Control group						
Study 1	No	Yes	No	No	No	No
Study 2	Yes	No	No	No	No	No
Study 3	No	Yes	Yes	No	No	No
Study 4	Yes	No	Yes	No	No	No
Study 5	No	Yes	Yes	No	Yes	No
Study 7	No	Yes	No	Yes	No	No
Study 8	Yes	No	No	Yes	No	No
Study 9	No	Yes	No	Yes	No	Yes
Experimental group						
Study 6	Yes	No	Yes	No	Yes	No
Study 10	Yes	No	No	Yes	No	Yes

The composition of each study is the result of the combination of three different methods—olfactory imprinting (trained or untrained worms), the use of cryoprotectant solutions (vitrification solution or slow freezing solution), and cryopreservation (vitrification or slow freezing).

the attractant benzaldehyde, applied by swiping 4  $\mu\text{L}$  of 1% benzaldehyde on the inside of the petri dish lid every hour for 8 hr. At the end of the 8-hr treatment period, the petri dish lid was replaced with a clean, unswiped lid. For untrained worms, the protocol was the same but we swiped 4  $\mu\text{L}$  of distilled water over the petri dish lid.

#### Slow freezing for *C. elegans*

Cryopreservation by slow freezing was performed 1 day after olfactory imprinting. Using the method of Brenner,<sup>2</sup> the worms at the L2 and L3 stages were introduced into a cryovial (cryogenic tube) with the traditional mix of cryoprotectant for slow freezing at 15% vol/vol glycerol in M9 Buffer (3 grams  $\text{KH}_2\text{PO}_4$ , 6 grams  $\text{Na}_2\text{HPO}_4$ , 5 grams  $\text{NaCl}$ , 1 mL of 1 M  $\text{MgSO}_4$ ,  $\text{H}_2\text{O}$  to 1 liter). The cryovials were transferred to a  $-80^\circ\text{C}$  freezer for 2 weeks. After 2 weeks, the worms were transferred to a petri dish with a lawn of *Escherichia coli* OP50.

#### SafeSpeed for *C. elegans*

One day after olfactory imprinting, the worms were vitrified. We used the SafeSpeed *C. elegans* protocol described in Barranco et al.<sup>14</sup> This method consists of two components: (1) SafeSpeed closed device (container for the worms), and (2) SafeSpeed closed system vitrification and warming media (cryoprotectant solutions). For each vitrification study using the SafeSpeed closed device, we vitrified five sets of worms at the L2 and L3 stages. The protocol started by transferring the worms from the petri dish to 50  $\mu\text{L}$  of Washing Solution (WS) for 2 min using a Pasteur pipette. The worms were then transferred to 100  $\mu\text{L}$  of Equilibration Solution (ES) for 10 min. Later, the worms were placed in 100  $\mu\text{L}$  of Vitrification Solution (VS) for 1 min, and then introduced into the SafeSpeed closed device. The device was closed with a heat-sealer to avoid contamination and then quickly immersed in liquid nitrogen. The device was held in liquid nitrogen for 30 min and rewarmed in a  $37^\circ\text{C}$  water bath. The SafeSpeed closed device was opened with scissors, and the worms were placed into

200  $\mu\text{L}$  of Thawing Solution (TS) at  $37^\circ\text{C}$  for 1 min and then quickly transferred to 200  $\mu\text{L}$  of the Diluent Solution (DS) for 3 min. To finish this protocol, the worms were washed twice in two drops of 100  $\mu\text{L}$  of WS, with 5 min for the first wash and 3 min for the second wash, and finally the worms were transferred to a petri dish with a lawn of *E. coli* OP50.

#### Chemotaxis assays

The odorant chemotaxis assays were performed on worms at the adult stage, 5 days after olfactory imprinting.<sup>9</sup> We used 12  $\times$  12-cm square low-salt agar plates (2% agar, 5 mM  $\text{KPO}_4$ , pH 6, 1 mM  $\text{MgCl}_2$ , and 1 mM  $\text{CaCl}_2$ ). On the outside bottom of each square dish, we drew eleven lines, marking off an area of 1  $\times$  12  $\text{cm}^2$  and assigned values from  $-6$  to  $+6$ . In the first delineated area of the plate (with value  $-6$ ), three drops ( $\mu\text{L}$  each) of 1 M sodium azide were placed in the agar with equal spacing. In the same area and in the same position as the drops of sodium azide, but in the lid of the square dish, three drops of  $\text{H}_2\text{O}_d$  (4  $\mu\text{L}$  each) were placed. On the other side of the square dish, in the area with value 6, the same drops (1  $\mu\text{L}$  each) of 1 M sodium azide were placed; however, the only difference was that on the lid of the dish we put three drops of 1% benzaldehyde (1/100 diluted in  $\text{H}_2\text{O}_d$ ) (see Supplementary Figure 1 in Remy and Hobert<sup>9</sup>).

We used a platinum wire to select and pick up individual worms from a petri dish with *E. coli* OP50, transferred the worms to a petri dish without food, and held them for 15 min. Then, 20 worms were transferred along the centerline of the square dish. The worms were counted every 15 min. At 1 hr, we had accumulated 80 values (each value designated the area where the worms were in that moment), and we calculated a migration report, which Remy and Hobert<sup>9</sup> referred to as the migration index (MI).

#### Statistical analysis

We used the software SPSS, version 20, to analyze the data set. The mean and the standard error of the variable MI

TABLE 2. NUMBER OF WORMS FOR EACH TYPE OF STUDY AND MIGRATION INDEX MEAN

S	NW	NCA	MI	SE	p
1	110	6	1.34	0.36	0.00
2	169	11	4.23	0.21	
3	115	7	2.00	0.38	
4	121	6	3.75	0.13	
5	122	8	1.62	0.15	
6	128	8	3.51	0.11	
7	108	6	1.51	0.23	
8	115	6	3.91	0.18	
9	114	6	1.73	0.25	
10	118	6	3.37	0.13	

Type of study (S), number of worms (NW), number of chemotaxis assay (NCA), migration index mean (MI), standard error (SE), and *p* value on the analysis of co-variance (ANOVA) test (*p*).

Types of studies (S) from 1 through 10: S1, untrained and not vitrified; S2, trained and not vitrified; S3, untrained, not vitrified and cryoprotectant solution; S4, trained, not vitrified and cryoprotectant solution; S5, untrained and vitrified; S6, trained and vitrified; S7, untrained, no slow freezing and cryoprotectant solution; S8, trained, no slow freezing and cryoprotectant solution; S9, untrained and slow freezing; and S10, trained and slow freezing.

were calculated for each type of study (Table 2), and to compare each study, different tests were applied over this variable (MI). The homogeneity of the variance was tested with the Levene test and the mean was compared with the one-factor analysis of variance (ANOVA) test. Finally, the Tahame test was used to test equality between pairs of study analyses.

Results

Cryopreservation of *C. elegans*: Slow freezing and SafeSpeed for *C. elegans*

The survival rates for our study did not show deviation from the expected original slow freezing method of Brenner<sup>2</sup> or the SafeSpeed method of Barranco.<sup>14</sup> The survival rate for slow freezing with L2–L3 worms was ~20%, and for vitrification was ~100%.

Chemotaxis assays and migration index

The trained worms (worms imprinted with benzaldehyde) preferred areas 5 and 6 on the square petri dish, close to the benzaldehyde drops, and the untrained worms preferred areas 1 and 2, demonstrating a native attraction to the benzaldehyde (Fig. 1). The mean of the MI was higher and very similar in all the studies with trained worms. The studies with untrained worms were also similar (Fig. 1). The highest value of the MI was 4.23 and was obtained with trained and unvitrified worms and the lowest value was 1.34 in the untrained and not vitrified worms (Table 2). In general, the response of the trained worms to the benzaldehyde was double that of the untrained worms, whether they were cryopreserved or not. The variance of MI did not show homogeneity (Levene=2.920, df1=9, df2=60, *p*=0.006) and the comparison of the media through one-factor ANOVA showed differences between groups (*F*=26.061, *p*=0.000). The Tahame test, for comparisons in pairs, showed differences between the two groups—trained and untrained worms—and no differences between the studies inside of each group.

There were no differences between trained and vitrified worms and trained and not vitrified worms (Tahame, *i-j*=0.72,

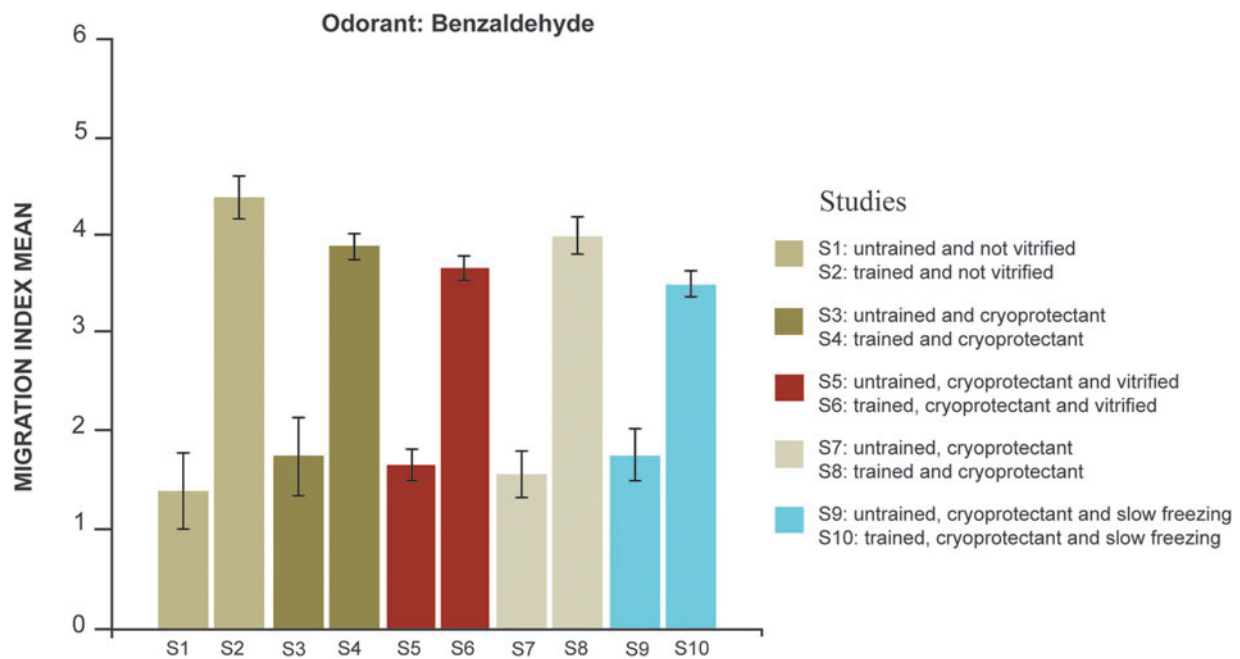


FIG. 1. Explanation of migration of untrained and trained worm studies: S1 (untrained and not vitrified) and S2 (trained and not vitrified); S3 (untrained and cryoprotectant) and S4 (trained and cryoprotectant); S5 (untrained, cryoprotectant and vitrified) and S6 (trained, cryoprotectant and vitrified); S7 (untrained and cryoprotectant) and S8 (trained and cryoprotectant); S9 (untrained, cryoprotectant and slow freezing) and S10 (trained, cryoprotectant and slow freezing).

$p=0.305$ ). Also, there were no differences between untrained and slow freezing and trained and slow freezing (Tahame,  $i-j=0.24$ ,  $p=0.138$ ). Both methods of cryopreservation did not show differences in the MI average of trained worms and were also similar to the MI of trained and worms that were not cryopreserved (Fig.1).

## Discussion

The study shows the first results related to the persistence of memory after vitrification. Prior to this study, there had been no other study or methodology to carry out this project or a similar project. The protocols that permitted us to examine memory in *C. elegans* were developed during the last 10 years,<sup>9,11,12</sup> and the protocols for successful vitrification of *C. elegans* with high survival rates were developed over the past few years.<sup>26,32</sup> *C. elegans* is one of the first animals that has been cryopreserved and revived by a completely artificial process<sup>2,26,32</sup> and is also one of the most important model organisms to be used in biology.

We used the *C. elegans* olfactory imprinting method of Remy and Hobert<sup>9</sup> in this study because it allows sufficient time to carry out all the steps in our cryopreservation protocol. The period between the olfactory imprinting (at the L1 stage) and the evaluation of memory in the MI assay (at the adult stage) is more than 26 hr. Specifically, the cryopreservation protocol in our study continues over 5 days. During this time, it is necessary to allow 30–45 min for vitrification processes, 15–30 min for slow freezing processes, and 24 hr for all processes to safely recover and verify the survival of the worms. Other methods<sup>12</sup> for testing memory in *C. elegans* where the time frame is not enough to allow for cryopreservation and recovery of the worms were discarded. Additions to our study might include testing the *ttx-3* and *sra-11* mutant strains, which are capable of responding to odorant cues but fail to be positively imprinted.<sup>9</sup> Alternatives to our study include possible future research, such as methods involving successful imprinting in *C. elegans* after four generations<sup>16</sup> as an epigenetic approach to memory, which would allow a new course to continue with our work in the future.

We demonstrated that cryoprotectants used in both the slow freezing and vitrification processes do not affect, alter, or change the mechanism that regulates the olfactory imprinting and long-term memory (Fig. 1). Second, we determined that the process of cryopreservation methods of slow freezing or vitrification do not affect, alter, or change this mechanism (Fig. 1). We also demonstrated that the results of the MI average obtained for the trained and untrained worms are similar to the results of the original authors of the odorant imprinting protocol.<sup>9</sup>

It is essential to emphasize the storage time in liquid nitrogen and the storage time in a  $-80^{\circ}\text{C}$  freezer. In the vitrification protocol, we maintained the worms for 30 min in liquid nitrogen before warming and for 2 weeks for slow freezing. For worms stored at  $-80^{\circ}\text{C}$  in the cryopreservation by slow freezing at the L1 stage, the time between olfactory imprinting and odorant chemotaxis assays was 5 days, although the worms were stored at  $-80^{\circ}\text{C}$  for 2 weeks. The worms have a life span of 15 days and develop from first larval stage (L1) to the mature adult stage in approximately 48 hr.<sup>5</sup>

There are several differences between the effects of vitrification and those of slow freezing in *C. elegans*. The most

important are the survival rate and the quality of the biological material cryopreserved. The method of vitrification by SafeSpeed for *C. elegans* offers a clear alternative to the traditional protocol by slow freezing due to the high survival rate (100% versus 25%–35%) and permits the cryopreservation of all stages of larvae, including the adult stage.<sup>31</sup> However, from the standpoint of preservation of the memory, the two protocols are equal. Although the value of the MI for trained worms that were cryopreserved by slow freezing (S10) was slightly lower than the MI obtained for trained and vitrified worms (S6) (Table 1), there was no significant difference between the studies.

Whether this result reflects the resilience of structural synaptic change or a more fragile chemical state change to the insult of cryopreservation remains unclear. More detailed chemotaxis assays of learning and memory in cryopreserved *C. elegans* are needed to determine whether all memory mechanisms are unaffected. Further research using methods for learning at different stages of *C. elegans* development to test memory retention after cryopreservation is needed. Additionally, further research on larger organisms with more complex nervous systems, such as the fruit fly *Drosophila melanogaster*, could prove to be beneficial to the issue of cryopreservation, including, specifically, memory retention after reviving.

## Conclusion

This study introduced the *C. elegans* nematode to olfactory imprinting for learning with the specific goal of testing the animal's memory retention after cryopreservation and revival. Our results show that the mechanisms that regulate odorant imprinting (a form of long-term memory) in young *C. elegans* have not been modified by either the process of vitrification or by slow freezing in the adult stage. This is the first evidence of preservation of memory after cryopreservation (vitrification or slow freezing).

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## Author Disclosure Statement

No conflicting financial interests exist.

## References

1. Brenner S. Nobel lecture. Nature's gift to science. *Biosci Rep* 2003;23:225–237.
2. Brenner S. The genetics of *Caenorhabditis elegans*. *Genetics* 1974;77:71–94.

3. Nass R, Miller III D, Blakely R. *C. elegans*: A novel pharmacogenetic model to study Parkinson's disease. *Parkinsonism Relat Disord* 2001;7:185–191.
4. Wu Y, Luo Y. Transgenic *C. elegans* as a model in Alzheimer's research. *Curr Alzheimer Res* 2005;2:37–45.
5. Riddle DL, Blumenthal T, Meyer B, Priess J. *C. elegans II*, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1997.
6. Rankin CH, Beck C, Chiba CM. *Caenorhabditis elegans*: A new model system for the study of learning and memory. *Behav Brain Res* 1990;37:89–92.
7. Mah KB, Rankin CH. An analysis of behavioral plasticity in male *Caenorhabditis elegans*. *Behav Neural Biol* 1992;58:211–221.
8. Dunn NA, Lockery SR, Pierce-Shinomura JT, Connery JT. A neural network model of chemotaxis predicts function of synaptic connections in the nematode *Caenorhabditis elegans*. *J Computational Neurosci* 2004;17:137–147.
9. Remy JJ, Hobert O. An interneuronal chemoreceptor required for olfactory imprinting in *C. elegans*. *Science* 2005;309:787–790.
10. Ardiel EL, Rankin CH. An elegant mind: Learning and memory in *Caenorhabditis elegans*. *Learn Mem* 2010;17:191–201.
11. Amano H, Maruyama IN. Aversive olfactory learning and associative long-term memory in *Caenorhabditis elegans*. *Learn Mem* 2011;18:654–665.
12. Kauffman A, Parsons L, Stein G, Wills A, Kaletsky R, Murphy C. *C. elegans* positive butanone learning, short-term, and long-term associative memory assays. *J Vis Exp* 2011;pii:2490.
13. Sasakura H, Mori I. Behavioral plasticity, learning, and memory in *C. elegans*. *Curr Opin Neurobiol* 2013;23:92–99.
14. Barranco D, Cabo V, Corral A, Risco R. Vitrification of *Caenorhabditis elegans* by ultra-fast warming rates. *Nat Methods* (in preparation).
15. Horn G. *Memory, Imprinting, and the Brain: An Inquiry into Mechanisms*. Clarendon Press, Oxford, 1985.
16. Remy JJ. Stable inheritance of an acquired behavior in *Caenorhabditis elegans*. *Curr Biol* 2010;20:877–878.
17. Mazur P. Freezing of living cells: Mechanisms and implications. *Am J Physiol* 1984;247(3 Pt 1):C125–C142.
18. Fahy GM, MacFarlane DR, Angell CA, Meryman HT. Vitrification as an approach to cryopreservation. *Cryobiology* 1984;21:407–426.
19. Risco R, Elmoazzen H, Doughty M, He X, Toner M. Thermal performance of quartz capillaries for vitrification. *Cryobiology* 2007;55:222–229.
20. Fahy GM, Wowk B, Pagotan R, Chang A, Phan J, Thomson B, Phan L. Physical and biological aspects of renal vitrification. *Organogenesis* 2009;5:167–175.
21. Vajta G, Kuwayama M. Improving cryopreservation systems. *Theriogenology* 2006;65:236–244.
22. Cobo A, Bellver J, Domingo J, Pérez S, Crespo J, Pellicer A, Remohí J. New options in assisted reproduction technology: The Cryotop method of oocyte vitrification. *Reprod Biomed Online* 2008;17:68–72.
23. Hamawaki A, Kuwayama M, Hamano S. Minimum volume cooling method for bovine blastocyst vitrification. *Theriogenology* 1999;51:165.
24. Vajta G, Holm P, Kuwayama M, Booth PJ, Jacobsen H, Greve T, Callesen H. Open pulled straw (OPS) vitrification: A new way to reduce cryoinjuries of bovine ova and embryos. *Mol Reprod Dev* 1998;51:53–58.
25. He X, Park, EYH, Flower, A, Yarmush ML, Toner, M. Vitrification by ultra-fast cooling at low concentration of cryoprotectants in a quartz microcapillary: A study using murine embryonic stem cells. *Cryobiology* 2008;56:223–232.
26. Risco R, Toner M. Quartz microcapillaries for *C. elegans* vitrification. *Cryobiology* 2008;57:320–321.
27. Seki S, Mazur P. Effect of warming rate on the survival of vitrified mouse oocytes and on the recrystallization of intracellular ice. *Biol Reprod* 2008;79:727–737.
28. Seki S, Mazur P. The dominance of warming rate over cooling rate in the survival of mouse oocytes subjected to a vitrification procedure. *Cryobiology* 2009;59:75–82.
29. Seki S, Mazur P. Ultra-rapid warming yields high survival of mouse oocytes cooled to –196 degrees C in dilutions of a standard vitrification solution. *PLoS One* 2012;7:e36058.
30. Mazur P, Seki S. Survival of mouse oocytes after being cooled in a vitrification solution to –196 degrees C at 95 degrees to 70,000 degrees C/min and warmed at 610 degrees to 118,000 degrees C/min: A new paradigm for cryopreservation by vitrification. *Cryobiology* 2011;62:1–7.
31. Risco R, Hebles M, Saa A, Vilches-Ferron MA, Sanchez-Martin P, Lucena E, Lucena M. Safespeed technology—the myth of ultrahigh cooling rates: A close device and a serum-free media for the vitrification human oocytes/embryos with the highest recovery rates. *European Society of Human Reproduction and Embryology 29th Annual Meeting* 2013;28:180–180.
32. Stiernagle T. Maintenance of *C. elegans*. In: Hope IA, ed. *C. elegans: A practical approach*. Oxford University Press, Oxford, 1999, pp. 51–67.

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